

REVIEW ARTICLE

Has nanotechnology led to improved therapeutic outcomes?

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Abstract

Nanoparticulate drug delivery systems provide new opportunities for solving issues associated with problematic drugs or disease states and have, therefore, created great expectations in the field of drug delivery. This review focuses on the potential benefits of nanoparticles compared with other conventional delivery systems. Several nanoparticulate drug delivery systems have already been marketed or are currently under development and are presented in this review. Results from clinical trials demonstrate that nanoparticulate formulations generally enable superior therapeutic outcomes than compared with standard formulations. Therefore, the implementation of nanotechnology in drug delivery represents an important advance with substantial potential to improve therapeutic effectiveness and increase patient's quality of life.

Keywords: nanotechnology, oral delivery, pulmonary delivery, parenteral delivery, poorly soluble drugs, efficacy of nanoparticles, safety of nanoparticles, drug targeting

Introduction

Over the past few decades, nanoparticulate drug delivery systems have become an area of extensive research as they enable bioavailability improvement of poorly water-soluble compounds as well as targeted delivery of active pharmaceutical ingredients to various tissues and organs^{1,2}. Generally, nanoparticles in drug delivery are defined as submicron colloidal particles ranging in size from 10 to 1000 nm³. The US Patent and Trademark Office (USPTO), however, defines nanotechnology using a scale from only 1 to 100 nm and slightly larger⁴. Owing to their reduced size, physical and chemical properties of nanoparticles differ significantly from their larger scale counterparts. The main difference is the significantly increased surface-area-to-volume ratio of nanoparticles than compared with microparticles. The enlarged surface area of nanoparticles directly translates into enhanced dissolution rates, which is especially beneficial in the delivery of poorly water-soluble drugs¹. Additionally, nanoparticles are characterized by increased biologic activity, for example, increased uptake and interaction with biological tissues, as a greater fraction of molecules is located at the surface allowing for more readily interaction with the external environment⁵. Due to their small

size, nanoparticles also differ from microparticles in terms of their *in vivo* distribution and ability to target specific tissues. For instance, nanoparticles of appropriate size and surface characteristics are able to extravasate through fenestrated tumor capillaries thereby passively accumulating in tumor tissue⁶. The preferential localization of nanoparticles at the site of interest is beneficial in that it reduces the occurrence of adverse side effects associated with nonspecific drug distribution. Overall, the unique features of nanoparticles have raised great expectations in the field of drug delivery because of their potential to improve clinical efficacy of problematic drug compounds. Compounds that previously could not be formulated due to insolubility or insufficient target specificity have the potential to be efficaciously delivered employing nanotechnology. Besides, nanoparticulate drug delivery systems offer exceptional flexibility as a wide variety of active agents including hydrophilic and hydrophobic drugs, proteins, vaccines, and biological macromolecules may be delivered via numerous routes of administration⁷. Few products have been commercialized yet, but several therapeutic agents are currently under development and expected to enter the market in the near future. It is the purpose of this review to highlight

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benefits of nanoparticulate formulations delivered via the oral, pulmonary, and parenteral route and to discuss the therapeutic outcomes achieved till date.

Nanoparticles delivered via the oral route

Benefits of nanoparticles for oral delivery

Dissolution velocity enhancement

When a drug is administered in form of a solid oral dosage form, it needs to first dissolve in the gastrointestinal (GI) fluids to then be absorbed through the intestinal mucosa into the bloodstream from where it is distributed throughout the body⁸. Consequently, the key factors that determine the rate and extent of oral drug absorption are the aqueous solubility of the drug and its permeability through the intestinal mucosa. The Biopharmaceutics Classification System (BCS) is a scientific framework that classifies drugs based on these two parameters into one of four categories: BCS I: high solubility, high permeability; BCS II: low solubility, high permeability; BCS III: high solubility, low permeability, and BCS IV: low solubility and low permeability⁹. A drug is considered highly permeable when the extent of absorption in humans is determined to be more than 90% of an administered dose, based on mass-balance or in comparison to an intravenous reference dose¹⁰. Furthermore, the criterion of high solubility is met, if the highest drug dose strength is soluble in 250 mL or less of aqueous media over the pH range of 1–7.5. It is estimated that about 60% of drugs that come directly from chemical synthesis are characterized by poor aqueous solubility (BCS II or BCS IV¹¹). While BCS IV compounds often do not enter later stages of drug development due to the unfavorable combination of low solubility and permeability, BCS II drugs may be successfully formulated employing several innovative formulation approaches aimed at improving their solubility/dissolution. Promising formulation strategies, which have emerged include particle size reduction, amorphization of crystalline drug compounds, self-emulsifying drug delivery systems, and solubilization via drug-cyclodextrin inclusion complexes¹². The conversion of a crystalline drug material into the amorphous form significantly increases the apparent solubility, but at the same time, it raises concerns regarding stability due to increased reactivity and hygroscopicity¹³. Self-emulsifying drug delivery systems, which are mixtures of oils and surfactants and sometimes co-solvents, can be employed for the formulation of highly lipophilic compounds. These systems spontaneously emulsify in the aqueous fluids of the GI tract to produce fine oil-in-water emulsions. Generally, the surfactant accounts for 30–60% (w/w) of the formulation, which increases the risk of undesirable side effects such as GI irritations¹⁴. The inclusion of poorly soluble drugs into the central cavity of cyclic oligosaccharides (cyclodextrins) is another potential way of increasing solubility. Still, cyclodextrins generally possess low drug inclusion capacity and in addition are limited to drugs that exhibit certain physico-chemical characteristics

that allow for complexation¹⁵. Particle size reduction as means of solubility enhancement has the advantage of being a nonspecific formulation strategy that it is applicable to almost any drug molecule. Exceptions include low-melting point drugs, which can potentially melt during size reduction due to high energy input. As particle size and specific surface area are inversely proportional, a size reduction will lead to an increase in surface area. For example, by reducing the particle size from 50 µm to 500 nm, the specific surface area will increase by a factor of 100. The increase in surface area will result in an enhanced dissolution velocity as described in the Noyes–Whitney equation (Eq. 1):

$$\frac{dM}{dt} = \frac{D \cdot A}{h} \cdot (c_s - c_i) \quad (1)$$

where dM/dt is the dissolution velocity, D the diffusion coefficient, A the surface area, h the diffusion layer thickness, and $(c_s - c_i)$ the concentration gradient between the diffusion layer and the bulk solution¹⁶. In addition, studies have shown that the size reduction also results in a decrease in the thickness of the diffusion layer surrounding each particle, a phenomenon that is especially pronounced for particles of less than 5 µm in size^{17,18}. According to Eq. 1, this will even further increase the dissolution rate. Lastly, the reduction in particle size also increases the saturation solubility. Generally, the saturation solubility is a constant depending on the compound, the dissolution medium, and the temperature. However, below a critical size of 1–2 µm, the saturation solubility becomes a function of particle size¹⁹. The saturation solubility (S) increases with decreasing particle radius (r) as described in the Ostwald–Freundlich equation (Eq. 2):

$$S = S_{\infty} \cdot \exp\left(\frac{2 \cdot \gamma \cdot M}{r \cdot \rho \cdot R \cdot T}\right) \quad (2)$$

where S_{∞} is the solubility of an infinitely large crystal, γ is the crystal-medium interfacial tension, M is the molecular weight of the solid, and ρ its density. According to the Noyes–Whitney equation, the dissolution velocity is further enhanced because it is directly proportional to the concentration gradient. On the basis of these facts, particle size reduction has become the method of choice for improving oral bioavailability of drugs that show dissolution-rate limited absorption.

Elimination of food effects

In the presence of food, drug absorption can generally be reduced, delayed, increased, or remain unchanged²⁰. Drugs that exhibit poor aqueous solubility commonly show an incomplete absorption as a result of their poor solubility in the GI fluids. Interestingly, when those drugs are taken with food an increase in absorption can be seen, which is referred to as a positive food effect. Gu and coworkers analyzed clinical data of food effects of 90 marketed compounds and found that 71% of BCS class II drugs showed a positive food effect²¹. This positive food

effect can be attributed to several factors including a delay in gastric emptying, pH alterations, and stimulation of bile secretion and portal blood flow²². Food effects can negatively influence therapeutic outcomes especially in the case of drugs that exhibit a narrow therapeutic window. If for example a high-fat meal is required to achieve effective drug levels, subtherapeutic concentrations may be obtained if the patient is taking the medicine without food²³. Nanoparticulate formulations are able to eliminate positive food effects of poorly soluble compounds and, therefore, have the potential of enhancing therapeutic outcomes by preventing the occurrence of either subtherapeutic or toxic concentrations of the drug¹.

Marketed products

Particle size reduction of drugs can generally be conducted by employing top-down or bottom-up approaches²⁴. Top-down processes consist of particle size reduction of large drug particles into smaller particles using various milling techniques, such as media milling or high-pressure homogenization techniques. In contrast, the bottom-up approach includes different particle formation processes, in which the drug is commonly dissolved in a suitable solvent and then precipitated on addition of an antisolvent in the presence of suitable stabilizers. A number of commercial drug products employing the top-down approach have been approved by the FDA. Specifically, the media milling technique, patented by ELAN under the name Nanocrystal[®], has shown great commercial success with already four oral drug products launched till date²⁵. Media milling is a type of wet milling, in which the milling chamber is charged with the milling media, the dispersion media, which can be aqueous or nonaqueous, the drug and suitable stabilizers²⁶. Particle sizes reported range from 80–400 nm and are a result of high shear forces generated by the impaction of the milling media with drug particles. The stabilizer(s) added to the milling chamber will adsorb to the nanocrystal surface thereby preventing agglomeration and improving wetting of particles. Typically, stable formulations are obtained at weight ratios of drug to stabilizer of 20:1–2:1. The final product after milling is a nanosuspension, which can be dried and particles further processed to obtain solid oral dosage forms such as tablets or capsules.

In the following paragraphs, all currently approved Nanocrystal[®]-based drug products for oral delivery will be discussed in more detail.

Rapamune[®] (Sirolimus)

Sirolimus, a triene macrolide antibiotic, is one of the most potent immunosuppressive agents for the prevention of graft rejection in organ transplantations²⁷. Still, the formulation of sirolimus represents a significant challenge because of the compound's low aqueous solubility of 2.6 µg/mL and high log *P* of greater than 5^{28,29}. In 1999, an oral lipid-based solution of sirolimus at a concentration of 1 mg/mL was approved by the FDA³⁰. Specifically, sirolimus is solubilized in a phosholipid-based carrier,

Phosal[®]50 PG, and polysorbate 80. The bioavailability of the oral solution is only 14%, which can be mainly attributed to metabolism of sirolimus by intestinal and hepatic cytochrome P 450 3A (CYP3A) enzymes and intestinal P-glycoprotein transport³¹. The influence of a high-fat meal on the systemic absorption of sirolimus after oral administration of the solution was investigated in 22 healthy volunteers³². It was found that sirolimus was absorbed more slowly when administered after a high-fat meal than when administered after fasting. The oral bioavailability was only increased to a modest extent of 35%, which was ascribed to a combination of enhanced absorption and inhibition of CYP3A and P-glycoprotein in the intestine. Besides low bioavailability, the solution of sirolimus has several other disadvantages such as unpleasant taste and need for refrigeration. To overcome problems associated with the oral solution, an oral tablet using the Nanocrystal[®] technology was developed and launched by Wyeth in 2000. This tablet provides patients with more convenient administration and storage than the oral solution. The pharmacokinetic parameters, efficacy, and safety of sirolimus tablet and oral solution for the prevention of renal allograft rejection were evaluated and compared in a multicenter, open-label study involving 447 patients³³. Pharmacokinetic analysis demonstrated that there were no statistically significant differences for the AUC_{0–24 h} and oral-dose clearance among the two formulations. It was further noted that intersubject variabilities for the AUC_{0–24 h} were large for both formulations. Besides, the *t*_{max} was significantly higher for the tablet when compared with the solution, indicating that sirolimus was more slowly absorbed from the tablet formulation. Furthermore, equivalence of both formulations was demonstrated regarding efficacy failure, defined as a composite of the first occurrence of biopsy-confirmed acute rejection, graft loss, or death in the first 3 months. In addition, no significant differences in patient and graft survival 3, 6, and 12 months after transplantation were observed between the treatment arms. No significant differences in the incidence of adverse events such as infections, malignancies, or death were seen between the tablet and solution treatment group. Overall, the results demonstrated that the sirolimus tablet formulation based on the Nanocrystal[®] technology is therapeutically equivalent to the oral solution formulation. The tablet formulation, however, provides additional benefits like stability at room temperature and improved palatability.

Emend[®] (Aprepitant)

Aprepitant is a highly selective antagonist of the neurokinin-1 receptor³⁴. Studies have shown that aprepitant combined with a 5-hydroxytryptamine₃ receptor antagonist and dexamethasone effectively prevents highly emetogenic chemotherapy-induced nausea and vomiting³⁵.

Aprepitant is a basic compound with a *pK_a* value of 9.7³⁶. The aqueous solubility of its thermodynamically stable polymorphic form I varies from 3 to 7 µg/mL in

the pH range of 2–10, while a log *P* value of 4.8 indicates high lipophilicity. During phase I and IIa clinical trials, significant positive food effects were seen with tablet formulations containing micronized aprepitant particles at a dose of 100 mg. Particularly, a threefold increase in exposure was observed under fed conditions. Data also suggested that high doses of aprepitant would be necessary to achieve efficacious drug levels. To guide formulation development, a beagle dog model was employed and the influence of particle size and food on the absorption of aprepitant was investigated³⁶. Specifically, suspensions of alpine-milled (mean particle size 5.49 µm), jet-milled (mean particle size 1.80 µm), wet-milled (mean particle size 0.48 µm), and Nanocrystal® media milled (mean particle size 0.12 µm) aprepitant were dosed orally. The decrease in particle size resulted in a significant increase in oral absorption: a fourfold increase in AUC was observed for the media-milled suspension compared with the alpine-milled suspension. Additionally, the alpine-milled suspensions exhibited a significant food effect, while a food effect was completely eliminated for the media-milled suspension. On the basis of the results, aprepitant was formulated using the Nanocrystal® technology. The nanosuspension obtained by media-milling is converted into beads by applying a colloidal coating dispersion in a Wurster-column coating process followed by filling into hard-gelatin capsules³⁷. The product was launched by Merck in April 2003 and is available in two dose strengths, 80 and 125 mg. Phase III clinical studies showed that the addition of aprepitant capsules to a standard regimen of ondansetron and dexamethasone improved the control of chemotherapy-induced nausea and vomiting associated with highly emetogenic cisplatin-based chemotherapy throughout the acute and delayed phases³⁵. In general, the aprepitant regimen was well tolerated, with adverse events comparable with those observed with the standard regimen. Overall, aprepitant can greatly enhance supportive care of patients with cancer. A study by Poli-Bigelli reported that 74.7% of patients showed minimal or no impact of chemotherapy-induced nausea and vomiting on daily life, compared with 63.5% for patients on standard therapy³⁸.

Tricor® (Fenofibrate)

Fenofibrate is a third-generation fibric acid derivative indicated for the treatment of hypercholesterolemia, combined dyslipidemia, remnant hyperlipidemia, hypertriglyceridemia, and mixed hyperlipemia³⁹. Several fenofibrate products have been marketed since its discovery in 1975, among them a micronized capsule formulation (TriCor®; 67 and 200 mg) in 1998 and a micro-coated tablet formulation (TriCor®; 54 and 160 mg) in 2001^{40,41}. The latter was developed to improve low fenofibrate bioavailability and high intersubject variability associated with the micronized capsule formulation. Particularly, a suspension containing micronised fenofibrate and the hydrophilic polymer polyvinylpyrrolidone was directly coated onto an inert hydrosoluble excipient core^{42,43}.

Even though, this tablet formulation demonstrated enhanced bioavailability combined with less variability in the maximum plasma concentration and extent of absorption between the fed and the fasted state, a positive food effect could not completely be eliminated⁴¹. In fact, bioavailability of the tablet formulation was increased by 35% when taken together with food compared with the fasted state⁴⁴. Therefore, fenofibrate tablets had to be taken with food. Since it cannot be expected that patients strictly adhere to the guidelines, inconsistent and suboptimal exposure is likely to occur with the tablet formulation. In November 2004, a new fenofibrate tablet formulation based on the Nanocrystal® technology was introduced in the United States to replace the micro-coated tablet formulation. The absorption of the new formulation (TriCor®; 48 and 145 mg) is not dependent on food and, therefore, results in more consistent blood concentrations. A study conducted by Maciejewski and Hilleman compared the effectiveness of the 145-mg nanocrystalline tablet formulation with the micro-coated 160-mg tablet formulation in patients with coronary heart disease and dyslipidemia⁴⁴. Patients were treated for a minimum of 6 months with fenofibrate 160 mg/day and were then switched to a minimum of 3-months treatment with fenofibrate 145 mg/day. Statistically significant reductions of 4.6% for mean triglycerides and 2.3% for low-density lipoprotein cholesterol (LDL) were observed after switching from fenofibrate 160 mg/day to fenofibrate 145 mg/day. Although no significant changes in the level of high-density lipoprotein cholesterol were noticed, the improvements in LDL and triglycerides indicate that nanoparticulate fenofibrate is more effective for treating combined dyslipidemia. Additionally, the elimination of a positive food effect for the nanoparticulate fenofibrate simplifies administration of the dosage form thereby greatly increasing patient compliance.

Megace®ES (Megestrol acetate)

Megestrol acetate is a synthetic derivative of naturally occurring progesterone⁴⁵. The drug is an effective appetite stimulant and, therefore, commonly prescribed to treat anorexia-cachexia syndrome associated with underlying diseases such as AIDS and cancer⁴⁶. Megestrol acetate is characterized as a BCS II drug with a low aqueous solubility of 2 µg/mL and high intestinal permeability⁴⁷. An oral suspension (Megace®) containing the drug in micronized form at a concentration of 40 mg/mL was first approved by the FDA in 1993. The initial adult dose recommended is 800 mg/mL requiring a large volume of 20 mL to be administered per day. In addition, the highly viscous character of the suspension results in relatively long residence times in the mouth or tubing. Therefore, the suspension is not well accepted by patients, particularly those depending on feeding tubes⁴⁸. Administration of megestrol acetate suspension also results in highly variable levels of systemic exposure. A pharmacokinetic study in patients infected with HIV who were administered a single oral dose of 800 mg for

21 days reported a high degree of interpatient variability in megestrol acetate pharmacokinetic parameters with an eightfold and fivefold range in the rate and extent of absorption, respectively⁴⁹. To increase bioavailability and decrease interpatient variability as well as suspension viscosity, an oral nanocrystal suspension was developed. Megace®ES, a more concentrated suspension containing 125 mg megestrol acetate per mL, was approved by the FDA in 2005. Due to the increase in drug concentration, the volume to be administered was reduced by a factor of four. Megace®ES is also 16 times less viscous (10 cP versus 163 cP) than the original Megace® formulation and, therefore, it is easier to consume, which will potentially increase patient compliance. The efficacy of Megace®ES oral suspension for treating anorexia-cachexia syndrome in patients with AIDS has been demonstrated in placebo-controlled, randomized efficacy trials. Megace®ES effectively stimulated appetite in nine of 10 patients and increased mean weight gain by 10.7 pounds versus placebo in 12 weeks⁵⁰.

In addition, a study conducted by Deschamps et al. investigated the pharmacokinetic properties of Megace®ES and Megace® under fed and fasted conditions in healthy male volunteers⁵¹. For the Megace®, oral suspension the AUC and C_{\max} in the fed state were increased by 52% and 86%, respectively, relative to the fasted state. In contrast, AUC and C_{\max} of Megace®ES in the fed state were only increased by 26% and 30%, respectively. Overall, AUC in the fed state was comparable for Megace® and Megace®ES. However, in the fasting state, Megace®ES resulted in significantly enhanced bioavailability. The results clearly indicate that Megace®ES is the preferred formulation for treating patients with anorexia-cachexia.

Nanoparticles delivered via the pulmonary route

Characteristics and benefits of nanoparticles for pulmonary delivery

The lung represents an attractive target for the delivery of drugs due to its unique anatomy and physiology. The respiratory system consists of two regions: the conducting airway, including trachea, bronchi, and bronchioles, and the respiratory region, comprising the respiratory bronchioles, alveolar ducts, and alveoli⁵².

The morphology and clearance mechanisms of these two regions differ considerably. While the conducting airways are characterized by a lower surface area and regional blood flow, the alveolar region of the lung features a large absorptive surface area and rich vascularization. Furthermore, clearance of foreign particles in the conducting airways is facilitated by entrapment in secreted mucus and transport toward the oropharynx by the continuous beating of the cilia of the respiratory epithelium (mucociliary escalator). In contrast, particles deposited in the alveolar region are primarily cleared from the lung by uptake into alveolar macrophages⁵³.

Inhalation of drugs through the respiratory tract can be utilized for both locally and systemically acting drugs. Local delivery of drugs to the lungs is attractive for many different diseases like asthma, pulmonary infections, or lung cancer as it directs the drug to the site of interest thereby achieving high local concentrations while minimizing systemic exposure. In addition, the alveolar region of the lung presents an excellent target for systemic delivery of drugs because of its large surface area of approximately 75–140 m²^{54,55}. Besides, the alveolar epithelium is extremely thin offering a very short airway-blood path that enables fast absorption. Pulmonary delivery of drug compounds also avoids first-pass metabolism and, therefore, offers the potential for higher systemic levels than would be achieved after oral delivery. Finally, pulmonary delivery represents a noninvasive delivery route for peptides and proteins that are normally administered via the parenteral route.

Drugs intended for local or systemic pulmonary delivery are generally administered in form of an aerosol that may be generated by employing metered dose inhalers, nebulizers, or dry powder inhalers. The deposition of aerosol particles within the respiratory tract is influenced by several factors including particle characteristics (e.g. particle size, density, hygroscopicity, shape, and electrical charge), the patient's breathing pattern (e.g. flow rate and ventilation volume), and lung morphology⁵⁶. Particle size is one of the most important characteristics as it influences both, the total amount of inhaled particles and the site of deposition in the respiratory tract. Generally, particles will deposit by three different mechanisms: inertial impaction, gravitational sedimentation, and Brownian diffusion⁵⁷. Impaction is the major deposition mechanism for particles with mass median aerodynamic diameters (MMAD) greater than 5 µm and primarily occurs in the upper airways and near-bronchial branching points⁵⁸. It can be further increased by high inspiratory flow rates. Particles deposited by inertial impaction will subsequently be swallowed and do, therefore, not provide any therapeutic response. Particles with MMADs between 1 and 5 µm will preferentially be deposited by gravitational sedimentation in small airways and respiratory bronchioles, while particles with MMADs below 500 nm are mostly confined to the alveoli. Particles of below 1 µm are, however, to the most part not deposited in the airways but rather exhaled after inspiration. This has led to a general MMAD recommendation of 1–3 µm for optimal deep lung delivery⁵⁹. Interestingly, ultrafine particles of less than 100 nm are also efficiently deposited in the deep lungs⁶⁰. A study conducted in healthy adults demonstrated high deposition rates of ultrafine particles with the total deposition fraction increasing with decreasing particle size from 100 nm to 40 nm and increasing respiratory time⁶⁰. However, the delivery of ultrafine particles is limited by several factors such as lack of appropriate formulation techniques and restriction of the dose that can be delivered⁶¹. To ensure efficient deposition of nanoparticles in

the deep lungs, “Trojan” particles have been suggested as an alternative⁶². Specifically, nanoparticles are formulated into larger porous structures either completely composed of nanoparticles or existing of nanoparticles and suitable carrier materials such as sugars or phospholipids. Once the micron-sized particles are effectively deposited in the lungs, they will disassociate to release the nanoparticles. The influence of different formulation parameters on the properties of these delivery systems has been widely explored; however, *in vivo* performance has yet to be evaluated^{63–65}.

Once particles have deposited in the respiratory system, they are subjected to clearance mechanisms. Particles that are deposited onto the surface of the ciliated upper bronchial area are normally removed by the mucociliary escalator within 24 hr⁶⁶. It has been reported that nanoparticles may overcome mucociliary clearance due to rapid displacement to the airway epithelium⁶⁷. In addition, particles deposited in the alveolar region are commonly removed through phagocytosis by alveolar macrophages. The process of phagocytosis is, however, dependent on a variety of factors including particle characteristics such as stiffness, coating, and size⁶⁸. Results from few studies indicate that uptake of inhaled nanoparticles by macrophages is insufficient and occurs rather sporadic and unintentional^{69–71}.

The combination of reduced phagocytotic and mucociliary clearance of nanoparticles provides the benefit of extended residence time of the drug at the site of action. In addition, nanoparticles facilitate a rapid onset of action, resulting from fast dissolution of the drug.

Potential applications of nanoparticles in pulmonary delivery

Pulmonary mycosis

Nanoparticles containing antifungals have been actively researched as inhaled therapeutic carriers for the prophylaxis and treatment of pulmonary mycoses. Aerosolized nanostructured formulations of itraconazole have shown promising results for the prevention of invasive pulmonary aspergillosis, a severe infection caused by *Aspergillus* spp⁷². Itraconazole is a poorly water-soluble compound that is commercially available for oral administration in form of capsules or an oral solution. The use of itraconazole is, however, challenged by the low and erratic bioavailability of the capsule formulation, considerable GI side effects associated with the oral solution, and numerous drug interactions due to systemic exposure⁷³. A study by Vaughn et al. compared the lung and serum concentrations in mice following oral and pulmonary dosing of amorphous nanoparticulate itraconazole compositions as well as the commercially available oral solution⁷⁴. High and sustained lung tissue concentrations were obtained after inhalation of nanoparticulate itraconazole. Particularly, pulmonary nanostructured itraconazole achieved significantly greater (>10-fold) lung tissue concentrations compared with the oral solution, while serum levels were maintained above the minimum lethal concentration of *Aspergillus fumigatus*. Furthermore, *in vivo*

efficacy in mice infected with *Aspergillus fumigatus* was demonstrated: nanostructured formulations of itraconazole significantly improved survival rate relative to the commercial itraconazole oral solution⁷². An additional study evaluated the safety of pulmonary nanostructured itraconazole in mice as assessed in changes in pulmonary histology. It was demonstrated that aerosolized administration of nanostructured itraconazole is safe with no evidence of bronchiolar, peribronchiolar, or perivascular inflammation observed in mouse lungs by histology⁷⁵.

Lung cancer

Nanoparticles have also been suggested for local lung delivery of anticancer drugs. The efficacy of chemotherapy in lung cancer is mostly insufficient due to rapid development of cancer cell resistance during treatment⁷⁶. As a consequence, higher doses of toxic anticancer drugs have to be administered, thereby increasing the risk of adverse side effects in other healthy tissues of the body. Localized delivery by inhalation increases accumulation in lung tumor cells and reduces systemic drug exposure and subsequent distribution into other organs. Nanoparticles are especially suitable for drug delivery to tumor cells since they selectively accumulate in the leaky tumor vasculature⁷⁷. Hitzman et al. determined the feasibility of delivering 5-fluorouracil in form of lipid-coated nanoparticles to hamsters for use in lung cancer chemotherapy⁷⁸. The drug is currently being administered intravenously with only a small fraction reaching the site of action. Nanoparticles investigated were coated with a mixture of tripalmitin and cetylalcohol to sustain drug release. Delivery of these particles via inhalation to hamsters resulted in efficacious and sustained concentrations of 5-fluorouracil in expected tumor sites. The drug serum level was nearly 1000 times lower than the level in the lung demonstrating the effectiveness of local delivery.

Local lung delivery of nanoparticles loaded with doxorubicin has also been studied⁷⁹. Particularly, doxorubicin nanoparticles were incorporated into inhalable lactose carrier particles using a spray-freeze-drying technique. Cytotoxic effects of particles were evaluated in lung cancer cell lines and compared with free doxorubicin. Doxorubicin nanoparticles demonstrated higher cytotoxicity compared with free drug possibly due to the fact that nanoparticles were readily internalized into cells by endocytosis, while free drug entered the cells through passive diffusion. Both, inhaled 5-fluorouracil and doxorubicin (Resmycin™) have been tested in form of aerosolized drug solutions in clinical trials in patients with malignant diseases in the lung^{80,81}. No clinical studies have been conducted involving nanoparticulate doxorubicin and 5-fluorouracil yet. However, *in vitro* data as well as *in vivo* data from animal models suggest that nanoparticulate systems may act superior to the free drug.

Asthma

Pulmonary delivery of budesonide in form of a nanocrystal suspension has been suggested for the treatment of

steroid-responsive pulmonary diseases such as asthma⁸². Budesonide is a poorly water-soluble compound that is available as a suspension for inhalation (Pulmicort Respules) with a nebulizer. The particle size of suspended particles is approximately 4.4 μm . The nanosuspension of budesonide that was suggested as an alternative was prepared employing the Nanocrystal[®] technology with particles in the range of 75–300 nm. The safety, delivery, and pharmacokinetics of nebulized nanobudesonide and Pulmicort Respules was evaluated in a randomized, double-blind, single-dose crossover study involving 16 healthy volunteers. Overall, both nebulized formulations were well tolerated. Nanobudesonide and Pulmicort Respules resulted in similar AUCs, suggesting a similar extent of pulmonary absorption. However, a higher C_{max} was seen with nanobudesonide and in addition the t_{max} was significantly smaller. Finally, nebulization of nanobudesonide resulted in significantly shorter nebulization times compared with Pulmicort Respules, which is advantageous with respect to patient compliance.

Pulmonary tuberculosis

Nanoparticles in pulmonary drug delivery can also be useful in targeting specific cell types such as alveolar macrophages⁸³. Uptake into these macrophages can be controlled by manipulating particle size and surface characteristics⁸⁴. Alveolar macrophage targeting using nanoparticulate carriers has been specifically investigated for the treatment of pulmonary tuberculosis. The infection is caused by *Mycobacterium tuberculosis*, which is spread via airborne dissemination. Once inhaled, the pathogen reaches the alveoli where it is taken up by alveolar macrophages⁸⁵. Pulmonary delivery of antitubercular drugs like isoniazid, rifampicin, and pyrazinamide ensures effective drug levels directly at the site of infection thereby decreasing systemic side effects and bypassing first-pass metabolism. The effectiveness of nanoparticulate pulmonary drug delivery systems in treating *Mycobacterium tuberculosis* has been demonstrated for different carrier materials in animal models. Pandey et al. investigated the suitability of poly (D,L-lactide-co-glycolide) (PLG) nanoparticles encapsulating isoniazid, rifampicin, and pyrazinamide for nebulization⁸⁶. The majority of drug-loaded PLG-nanoparticles were in the size range of 186–290 nm, while the MMAD of nebulized particles was determined to be 1.88 μm favoring broncho-alveolar lung delivery. Single nebulization to guinea pigs resulted in sustained therapeutic drug levels in plasma and lungs. Relative bioavailability compared with oral administration was enhanced by factor 12.7, 32.8, and 14.7 for rifampicin, isoniazid, and pyrazinamide, respectively. Upon pulmonary administration of drug-loaded nanoparticles to guinea pigs infected with *Mycobacterium tuberculosis*, no tubercle bacilli were detected in the lungs after five doses of treatment. In contrast, 46 daily doses of orally administered drug were required to obtain an equivalent therapeutic benefit. A study conducted by Sharma et al. investigated the influence of a lectin coating on the dose

frequency of antitubercular drugs encapsulated into PLG-nanoparticles⁸⁷. The lectin coating was chosen based on the assumption that interactions of lectin with its receptors on the alveolar epithelium could potentially sustain drug release. In fact, after pulmonary administration of coated and uncoated PLG-nanoparticles, rifampicin was detectable in plasma for 13–14 and 4–6 days, respectively, indicating effective binding of lectin to the alveolar epithelium. Besides biodegradable polymers like PLGA and PLG, lipid materials have been investigated as nanocarrier materials for treatment of pulmonary tuberculosis. Particularly, solid lipid nanoparticles (SLN) have shown promising therapeutic potential⁸⁸. SLN exhibit several advantages over polymeric nanoparticles such as higher tolerability and faster *in vivo* degradation^{89,90}. However, research related to pulmonary delivery of SLN is still in its infancy and their full potential has yet to be explored⁹¹. Overall, nanoparticulate carriers for pulmonary delivery of antitubercular drugs have demonstrated the potential to effectively target alveolar macrophages thereby minimizing toxic side effects and dosing frequency. These, in turn, are key factors for improving therapeutic outcomes as well as patient compliance.

Insulin-dependent diabetes mellitus

Nanoparticles for systemic delivery have the potential to sustain drug release in the lung and accordingly systemic circulation as shown for insulin. Kawashima and coworkers prepared biodegradable PLGA-nanospheres with mean diameters of 400 nm by a modified emulsion solvent diffusion method⁹². An aqueous dispersion of insulin-loaded-PLGA nanospheres and an insulin solution serving as a control were nebulized into the trachea of fasted guinea pigs for 20 min. The PLGA-nanosphere dispersion significantly reduced blood glucose levels over a prolonged period of 48 hr compared with only 6 hr for the insulin solution. In addition, Zhang et al. investigated the suitability of insulin-loaded polybutylcyanoacrylate nanoparticles for systemic lung delivery⁹³. The nanoparticle dispersion was given intratracheally to rats and their hypoglycemic effect compared with an insulin solution administered either intratracheally or subcutaneously by bolus injection. It was shown that the hypoglycemic effect of insulin-loaded nanoparticles lasted much longer than in the case of the insulin solution. However, results indicated that bioavailability of insulin-loaded nanoparticles by intratracheal delivery was lower than by subcutaneous administration, with a relative bioavailability of insulin-loaded nanoparticles of 57.2% compared with subcutaneous administration of the solution.

In both cases, the polymeric nanocarrier materials sustained insulin release reducing glucose blood levels over a prolonged time compared with a pulmonary-delivered insulin solution. Overall, pulmonary delivery of insulin nanoparticles has the potential to increase patient compliance by reducing dose frequency. Additionally, it represents a more patient-friendly noninvasive form of administration.

Concerns regarding nanoparticles in pulmonary delivery

The high surface area of nanoparticles, which makes them so attractive for drug delivery, also raises concerns regarding potential toxicity since it is chemically more reactive. As the respiratory tract is the primary entrance way for nanoparticles into the human body, numerous studies have studied the potential of nanosized-airborne pollutants to cause adverse health effects. These studies have reported the occurrence of inflammatory reactions as well as carcinogenicity^{94,95}. Early studies in rats showed that ultrafine TiO₂ particles (below 50 nm) have a significantly greater pulmonary inflammatory potency than larger TiO₂ particles (~200 nm⁹⁶). Data suggest that ultrafine particles due to their high surface area lead to oxidative stress and calcium changes in alveolar macrophages and epithelial cells thereby activating cells for inflammation^{97,98}. Furthermore, studies have shown that poorly soluble inhaled nanoparticles increase the incidence of lung tumors in rats^{99,100}. It was hypothesized that inflammation and inflammatory cell-derived oxidants as induced by high doses of insoluble nanoparticles initiate and promote neoplastic transformation of epithelial cells¹⁰¹. However, it is important to note that most studies have investigated inorganic and insoluble materials like TiO₂, while carrier materials for drug delivery are commonly of soluble or biodegradable and organic nature. On the basis of that, physiological responses caused by therapeutic materials are expected to notably differ from those originated by insoluble materials⁸³. Concerns regarding the toxicity of nanoparticles also stem from the fact that they can be taken up by the epithelium gaining access to the systemic circulation and extrapulmonary organs causing undesirable side effects¹⁰². Overall, the influence of key factors such as size, persistence, solubility, and charge has to be further investigated to understand nanoparticles full impact on the body.

Nanoparticles delivered via the intravenous route

Benefits of nanoparticles delivered intravenously

In the case of intravenous administration, the medication is directly injected into a patient's vein by means of a small-volume bolus injection (<50 mL) or an infusion (50–1000 mL¹⁰³). Since the drug is directly delivered to the bloodstream, a rapid onset of action is achieved, which is often vital in emergency situations such as cardiac arrest or status epilepticus¹⁰⁴. Besides, intravenous administration may be employed for drugs that show poor absorption from the GI tract or undergo extensive first-pass metabolism after oral administration.

To ensure safe intravenous therapy, formulations have to be sterile and free from pyrogens and visible particulate matter¹⁰⁵. In addition, pH and tonicity of intravenous formulations should preferably be matched to physiological values to avoid pain and tissue irritation. As water is the principal constituent of blood, intravenous formulations are ideally formulated as aqueous solutions to maximize

in vivo tolerability. However, poorly water-soluble drugs intended for intravenous administration often need to be solubilized with large amounts of organic cosolvents and surfactants or cyclodextrins, which may cause undesirable side effects¹. Furthermore, dilution of the solubilized formulation in the bloodstream may result in precipitation of the drug, which in turn may induce inflammation of the vein wall¹⁰⁶.

Intravenous delivery of poorly water-soluble drugs in form of aqueous suspensions is a viable alternative that avoids the use of harsh solubilizing excipients. The formulation of suspensions for intravenous administration, however, requires the particle size to be reduced to the submicron range (nanosuspensions) to prevent vascular occlusion¹⁰⁷. The use of drug nanosuspensions also ensures conformance with the limits specified by the USP regarding particulate matter in injections, which requires that the average number of particles present in the units tested does not exceed 3000 per container equal to or greater than 10 µm and 300 per container equal to or greater than 25 µm when the microscopic particle count test is applied¹⁰⁸. As pointed out earlier, the use of nanosuspensions is also beneficial in that it goes along with a substantial increase in the dissolution rate. Besides, nanosuspensions are favored when low-potency drugs are formulated since they offer higher mass per volume loadings¹⁰⁹. Also, nanosuspensions are generally preferred over solutions as drugs are more chemically stable in the solid-state than in the dissolved-state. Finally, administering drugs in form of nanoparticles enables targeted delivery due to their particulate nature and the possibility of surface modification. Targeted delivery of nanoparticles via intravenous administration has been especially explored to treat various types of cancer.

Nanoparticles in cancer therapy

Size and surface characteristics

The size and surface characteristics of intravenously administered nanoparticles are key factors determining their circulation time and target specificity. To stay in the systemic circulation for a sufficient time, nanoparticles need to escape the reticuloendothelial system (RES), which is predominantly distributed in liver, lung, spleen, and bone marrow¹¹⁰. Surface modification with water-soluble polymers such as PEG has shown to reduce RES uptake of nanoparticles by minimizing interactions between nanoparticles and opsonin molecules¹¹¹. In addition, smaller particles (~100 nm) escape the RES more easily. The tumor vasculature is known to be leaky with junctions between the epithelial cells ranging from 100 to 600 nm depending on the type of tumor¹¹². Therefore, the optimal size of nanoparticles is thought to be between 10 and 100 nm.

Passive targeting

Nanoparticles that exhibit the size and surface features described above will passively accumulate at the tumor site. Accumulation in healthy tissue is theoretically

restricted by the fact that normal vasculature has much tighter junctions of less than 10 nm¹¹⁰. Besides increased permeability, tumor vessels are also characterized by a less effective lymphatic drainage system so that drugs are retained in the interstitium for prolonged times.

Active targeting

Nanoparticles offer the additional benefit that their surface can be easily modified, for example, through the addition of targeting ligands¹¹³. Targeting ligands in cancer research commonly include peptides, proteins, and antibodies all of which are added to facilitate binding to receptors that are over-expressed in tumor cells. Since they enable selective uptake into tumor cells, they have become an active area of research. Promising results have been demonstrated with biodegradable poly (d,l-lactic-co-glycolic acid)-block-poly (ethylene glycol) (PLGA-b-PEG) nanoparticles loaded with doxorubicin that were surface-targeted to the extracellular domain of a prostate-specific membrane antigen by conjugation of an RNA aptamer¹¹⁴. *In vitro* cytotoxicity tested in prostate epithelial cells demonstrated that nanoparticles carrying the RNA aptamer were significantly more cytotoxic as compared with control nanoparticles that were missing the targeting ligand. The aptamer-nanoparticles also exhibited remarkable efficacy and reduced toxicity in xenograft mice. After a single intratumoral injection, complete tumor reduction was observed in five of seven mice and 100% of animals survived the 109-day study. In contrast, only two of seven mice that were administered nanoparticles without the targeting aptamer showed complete tumor reduction and only 57% survived the study.

In vivo biodistribution studies comparing targeted and nontargeted nanoparticles in a murine model demonstrated that the enhanced efficacy of targeted nanoparticles is not necessarily a result of increased tumor accumulation¹¹⁵. More generally, targeting ligands increase cellular internalization without affecting overall tumor uptake. This highlights the importance of the physicochemical properties of the nanoparticle carrier material itself as it will largely determine the overall biodistribution and pharmacokinetics of the delivery system.

Multidrug resistance

Resistance of tumor cells to a variety of anticancer drugs, for example, through P-glycoprotein-mediated efflux, represents a major challenge in achieving clinical efficacy. Results from different studies suggest that nanoparticulate delivery systems are able to circumvent efflux through transmembrane P-glycoprotein^{116,117}. One suggestion is that since nanoparticles enter the cell via endocytosis, they avoid recognition by P-glycoprotein reaching high intracellular concentrations. However, there is no ultimate verification that nanoparticles are able to evade P-glycoprotein-mediated efflux in humans, even though clinical data points to it¹¹³.

Marketed product

Abraxane® (Nanoparticle albumin-bound (nab™)-paclitaxel)

Paclitaxel is a mitotic inhibitor that has a wide spectrum of antitumor activity when used as monotherapy or in combination chemotherapy regimens¹¹⁸. The poorly water-soluble molecule was first formulated in form of a nonaqueous solution for intravenous infusion (Taxol®). In this solution, paclitaxel is solubilized in a mixture of polyoxyethylated castor oil (Cremphor®EL) and ethanol. The formulation can cause considerable side effects, particularly neuropathy and hypersensitivity reactions, both of which are linked to the use of Cremphor®EL. To prevent the occurrence of hypersensitivity reactions, pre-medication with steroids and antihistamines is required. To overcome the problems associated with the solution, a nanoparticulate paclitaxel formulation (Abraxane®) was developed. The nanoparticles are composed of a hydrophobic drug core surrounded by a hydrophilic albumin coating. The particles are manufactured by forming an oil-in-water emulsion of a water-immiscible solvent containing paclitaxel and an aqueous phase containing albumin. This emulsion is high-pressure homogenized to reduce the particle size and cross-link the albumin to stabilize particles¹¹⁹. Finally, the solvent is evaporated to yield a colloidal dispersion containing albumin-coated nanoparticles of mean diameters of 130 nm. The small particle size allows for sterile filtration, an important feature, since the dispersion cannot be sterilized by conventional means such as autoclaving due to the high amount of albumin present. In addition, albumin may serve as a substrate for microbial growth; therefore, the formulation is stored in lyophilized form and has to be reconstituted before infusion¹²⁰. Particularly, nanoparticle albumin-bound paclitaxel (nab-paclitaxel) is reconstituted in normal saline at concentrations of 2–10 mg/mL compared with 0.3–1.2 mg/mL for the Cremphor®EL containing solution. Therefore, the volume to be administered and consequently the infusion time could be significantly reduced¹¹⁸.

Nab-paclitaxel can leave the circulation through leaky tumor microvasculature and accumulate in the interstitium. In addition, the albumin shell facilitates interaction with receptors on endothelial cells (gp60) and in the tumor interstitium (SPARC). Specifically, albumin-gp60 binding leads to endothelial transcytosis of nab-paclitaxel into the extravascular space, where SPARC, which is overexpressed in many types of cancer, entraps the albumin in tumor cells resulting in high intratumoral accumulation¹²¹. In fact, mice bearing human tumor xenografts treated with nab-paclitaxel showed a paclitaxel area under the curve that was 33% higher than for Cremphor®EL-based paclitaxel, indicating more effective intratumoral accumulation¹²². It was also determined that the maximum-tolerated dose is 300 mg/m², which is about 70% higher than for the Cremphor®EL-based solution of paclitaxel.

Furthermore, phase I clinical trials demonstrated that paclitaxel C_{max} and area under the curve values

increase linearly over a dose range of 135–300 mg/m² ¹¹⁸. This represents a huge advantage over the Cremophor®EL-paclitaxel formulation, which exhibits parabolic pharmacokinetics causing an unpredictable relationship between dose, efficacy, and risk of toxicity¹²³. The nonlinear pharmacokinetics has been related to the formation of Cremophor®EL micelles in the plasma, which entrap paclitaxel, sequestering it in the plasma¹²⁴.

A phase III clinical trial involving 454 patients with metastatic breast cancer demonstrated that nab-paclitaxel administered at a dose of 260 mg/m² resulted in significantly higher response rates (33%) compared with CremophorEL®-based paclitaxel administered at a dose of 175 mg/m² (19%¹²⁵). Despite the higher dose of nab-paclitaxel that was delivered, patients treated experienced significantly less neutropenia than patients treated with CremophorEL®-based paclitaxel. The incidence of hypersensitivity reactions (any grade) was low for nab-paclitaxel (<1%) despite the absence of premedication.

In summary, nab-paclitaxel overcomes several limitations of the Cremophor®EL-based paclitaxel. It clearly represents an important advance in the treatment of patients with metastatic breast cancer offering significantly improved efficacy without compromising patient safety and quality of life.

Conclusions

Nanoparticulate drug delivery has clearly provided new opportunities for solving issues associated with problematic drugs or disease states. Great commercial success has been achieved in the area of oral drug delivery either by formulation of new water-insoluble drugs or reformulation of established therapeutic agents. Results from clinical trials demonstrate that nanoparticulate formulations generally enable superior therapeutic outcomes than compared with standard formulations. Even in the case where nanoparticulate drug delivery systems have shown no superiority but rather equivalence regarding their efficacy; they commonly offer additional benefits such as improved stability or simplified administration thereby improving patient compliance and consequently therapeutic responses.

Despite significant accomplishments in the area of nanoparticulate drug delivery, some challenges still persist. Particularly, concerns regarding the safety of nanoparticles represent a considerable barrier toward successful establishment of nanoparticulates in the field of drug delivery. To make nanoparticulate drug delivery systems available for a wide variety of disease states and delivery routes, their full biological impact on the human body has to be more closely explored.

Overall, the implementation of nanotechnology in drug delivery represents an important advance with substantial potential to improve therapeutic outcomes and increase patient's quality of life.

Declaration of interest

The authors report no conflict of interest.

References

1. Merisko-Liversidge EM, Liversidge GG. (2008). Drug nanoparticles: formulating poorly water-soluble compounds. *Toxicol Pathol*, 36:43–48.
2. Yih TC, Al-Fandi M. (2006). Engineered nanoparticles as precise drug delivery systems. *J Cell Biochem*, 97:1184–1190.
3. Rao GC, Kumar MS, Mathivanan N, Rao ME. (2004). Nanosuspensions as the most promising approach in nanoparticulate drug delivery systems. *Pharmazie*, 59:5–9.
4. United States Patent and Trademark Office. (2010). Nanotechnology-Class Definition. [Online] Available at: <http://www.uspto.gov/web/patents/classification/uspc977/defs977.htm>. Accessed on 29 October 2010.
5. Oberdörster G, Oberdörster E, Oberdörster J. (2005). Nanotoxicology: an emerging discipline evolving from studies of ultrafine particles. *Environ Health Perspect*, 113:823–839.
6. Ferrari M. (2005). Cancer nanotechnology: opportunities and challenges. *Nat Rev Cancer*, 5:161–171.
7. Hans ML, Lowman AM. (2002). Biodegradable nanoparticles for drug delivery and targeting. *Curr Opin Solid State Mat Sci* 6:319–327.
8. Dunne A, Devane J, O'Hara T. (1999). The relationship between in vitro drug dissolution and in vivo absorption. *J R Stat Soc Ser D-Stat* 48:125–133.
9. Amidon GL, Lennernäs H, Shah VP, Crison JR. (1995). A theoretical basis for a biopharmaceutical drug classification: the correlation of *in vitro* drug product dissolution and *in vivo* bioavailability. *Pharm Res*, 12:413–420.
10. FDA. (2000). Guidance for industry: waiver of in vivo bioavailability and bioequivalence studies for immediate release dosage forms based on a biopharmaceutical classification system. Center for Drug Evaluation and Research.
11. Muller RH, Junghanns JAH. (2006). Drug nanocrystals/nanosuspensions for the delivery of poorly soluble drugs. In: Tochilin V (Ed.) *Nanoparticulates as drug carriers*. Imperial College Press, London, UK. 307–328.
12. Fahr A, Liu X. (2007). Drug delivery strategies for poorly water-soluble drugs. *Expert Opin Drug Deliv*, 4:403–416.
13. Hilden LR, Morris KR. (2004). Physics of amorphous solids. *J Pharm Sci*, 93:3–12.
14. Tang B, Cheng G, Gu JC, Xu CH. (2008). Development of solid self-emulsifying drug delivery systems: preparation techniques and dosage forms. *Drug Discov Today*, 13:606–612.
15. Huang L, Dong J. (2008). Formulation strategies and practice used for candidates with water-insoluble properties for toxicology, biology, and pharmacology studies in discovery support. In: Liu R (Ed.) *Water-insoluble drug formulation*. CPR Press, Boca Raton, USA. 113–132.
16. Noyes A, Whitney W. (1897). The rate of solution of solid substances in their own solutions, *J Am Chem Soc* 19: 930–934.
17. Bisrat M, Nystrom C. (1988). Physicochemical aspects of drug release. 8. the relation between particle-size and surface specific dissolution rate in agitated suspensions. *Int J Pharm* 47:223–231.
18. Mosharraf M, Nystrom C. (1995). The effect of particle-size and shape on the surface specific dissolution rate of micro-sized practically insoluble drugs. *Int J Pharm* 122:35–47.
19. Junghanns JU, Müller RH. (2008). Nanocrystal technology, drug delivery and clinical applications. *Int J Nanomedicine*, 3:295–309.
20. Welling PG. (1977). Influence of food and diet on gastrointestinal drug absorption: a review. *J Pharmacokinet Biopharm*, 5:291–334.
21. Gu CH, Li H, Levons J, Lentz K, Gandhi RB, Raghavan K et al. (2007). Predicting effect of food on extent of drug absorption based on physicochemical properties. *Pharm Res*, 24:1118–1130.

22. Welling PG. (1996). Effects of food on drug absorption. *Annu Rev Nutr*, 16:383–415.
23. Lentz KA. (2008). Current methods for predicting human food effect. *AAPS J*, 10:282–288.
24. Keck CM, Müller RH. (2006). Drug nanocrystals of poorly soluble drugs produced by high pressure homogenisation. *Eur J Pharm Biopharm*, 62:3–16.
25. Elan Drug Technologies. (2010). Nanocrystal® Technology. Commercialized Products. [Online] Available at: http://www.elandrugtechnologies.com/nanocrystal_technology/commercialised. Accessed on 17 October 2010.
26. Merisko-Liversidge E, Liversidge GG, Cooper ER. (2003). Nanosizing: a formulation approach for poorly-water-soluble compounds. *Eur J Pharm Sci*, 18:113–120.
27. Vazquez EM. (2000). Sirolimus: A new agent for prevention of renal allograft rejection. *Am J Health-Syst Pharm* 57:437–448.
28. Simamora P, Alvarez JM, Yalkowsky SH. (2001). Solubilization of rapamycin. *Int J Pharm*, 213:25–29.
29. Gandhi PJ, Murthy ZVP. (2010). Kinetic study of ultrasonic antisolvent crystallization of sirolimus. *Cryst Res Technol* 45: 321–327.
30. Shen LJ, Wu FL. (2007). Nanomedicines in renal transplant rejection–focus on sirolimus. *Int J Nanomedicine*, 2:25–32.
31. Lampen A, Zhang Y, Hackbarth I, Benet LZ, Sewing KE, Christians U. (1998). Metabolism and transport of the macrolide immunosuppressant sirolimus in the small intestine. *J Pharmacol Exp Ther*, 285:1104–1112.
32. Zimmerman JJ, Ferron GM, Lim HK, Parker V. (1999). The effect of a high-fat meal on the oral bioavailability of the immunosuppressant sirolimus (rapamycin). *J Clin Pharmacol*, 39:1155–1161.
33. Mathew TH, Van Buren C, Kahan BD, Butt K, Hariharan S, Zimmerman JJ. (2006). A comparative study of sirolimus tablet versus oral solution for prophylaxis of acute renal allograft rejection. *J Clin Pharmacol*, 46:76–87.
34. Majumdar AK, Howard L, Goldberg MR, Hickey L, Constanzer M, Rothenberg PL et al. (2006). Pharmacokinetics of aprepitant after single and multiple oral doses in healthy volunteers. *J Clin Pharmacol*, 46:291–300.
35. Hesketh PJ, Grunberg SM, Gralla RJ, Warr DG, Roila F, de Wit R et al.; Aprepitant Protocol 052 Study Group. (2003). The oral neurokinin-1 antagonist aprepitant for the prevention of chemotherapy-induced nausea and vomiting: a multinational, randomized, double-blind, placebo-controlled trial in patients receiving high-dose cisplatin–the Aprepitant Protocol 052 Study Group. *J Clin Oncol*, 21:4112–4119.
36. Wu Y, Loper A, Landis E, Hettrick L, Novak L, Lynn K et al. (2004). The role of biopharmaceutics in the development of a clinical nanoparticle formulation of MK-0869: a Beagle dog model predicts improved bioavailability and diminished food effect on absorption in human. *Int J Pharm*, 285:135–146.
37. Olver I, Shelukar S, Thompson KC. (2007). Nanomedicines in the treatment of emesis during chemotherapy: focus on aprepitant. *Int J Nanomedicine*, 2:13–18.
38. Poli-Bigelli S, Rodrigues-Pereira J, Carides AD, Julie Ma G, Eldridge K, Hipple A et al.; Aprepitant Protocol 054 Study Group. (2003). Addition of the neurokinin 1 receptor antagonist aprepitant to standard antiemetic therapy improves control of chemotherapy-induced nausea and vomiting. Results from a randomized, double-blind, placebo-controlled trial in Latin America. *Cancer*, 97:3090–3098.
39. Tziomalos K, Athyros VG. (2006). Fenofibrate: a novel formulation (Triglide) in the treatment of lipid disorders: a review. *Int J Nanomedicine*, 1:129–147.
40. Guay DR. (1999). Micronized fenofibrate: a new fibric acid hypolipidemic agent. *Ann Pharmacother*, 33:1083–1103.
41. Guichard JP, Blouquin P, Qing Y. (2000). A new formulation of fenofibrate: suprabioavailable tablets. *Curr Med Res Opin*, 16:134–138.
42. Keating GM, Ormrod D. (2002). Micronised fenofibrate: an updated review of its clinical efficacy in the management of dyslipidaemia. *Drugs*, 62:1909–1944.
43. Stamm A, Seth P. (2003). Fenofibrate pharmaceutical composition having high bioavailability and method for preparing it. US Patent No. 6589552.
44. Maciejewski S, Hilleman D. (2008). Effectiveness of a fenofibrate 145-mg nanoparticle tablet formulation compared with the standard 160-mg tablet in patients with coronary heart disease and dyslipidemia. *Pharmacotherapy*, 28:570–575.
45. Femia RA, Goyette RE. (2005). The science of megestrol acetate delivery: potential to improve outcomes in cachexia. *Biodrugs*, 19:179–187.
46. Berenstein EG, Ortiz Z. (2005). Megestrol acetate for the treatment of anorexia-cachexia syndrome. *Cochrane Database Syst Rev*, CD004310.
47. Zhang ZB, Shen ZG, Wang JX, Zhao H, Chen JF, Yun J. (2009). Nanonization of megestrol acetate by liquid precipitation. *Ind Eng Chem Res* 48:8493–8499.
48. Hovey D, Pruitt J, Ryde T. (2005). Nanoparticulate megestrol formulations. US Patent Application. Pub.No. US 2005/0233001 A1.
49. Graham KK, Mikolich DJ, Fisher AE, Posner MR, Dudley MN. (1994). Pharmacologic evaluation of megestrol acetate oral suspension in cachectic AIDS patients. *J Acquir Immune Defic Syndr*, 7:580–586.
50. Par Pharmaceuticals. Par Pharmaceuticals Announces FDA Approval of Megace ES for Anorexia, Cachexia, or an Unexplained, Significant Weight Loss in Patients with a Diagnosis of AIDS. [Online] Available at: <http://www.parpharm.com/generics/index>. Accessed on 19 October 2010.
51. Deschamps B, Musaji N, Gillespie JA. (2009). Food effect on the bioavailability of two distinct formulations of megestrol acetate oral suspension. *Int J Nanomedicine*, 4:185–192.
52. Agu RU, Ugwoke MI, Armand M, Kinget R, Verbeke N. (2001). The lung as a route for systemic delivery of therapeutic proteins and peptides. *Respir Res*, 2:198–209.
53. Suarez S, Hickey AJ. (2000). Drug properties affecting aerosol behavior. *Respir Care*, 45:652–666.
54. Gehr P, Bachofen M, Weibel ER. (1978). The normal human lung: ultrastructure and morphometric estimation of diffusion capacity. *Respir Physiol*, 32:121–140.
55. Laube BL. (2005). The expanding role of aerosols in systemic drug delivery, gene therapy, and vaccination. *Respir Care*, 50:1161–1176.
56. Sweeney TD, Brain JD. (1991). Pulmonary deposition: determinants and measurement techniques. *Toxicol Pathol*, 19:384–397.
57. Courrier HM, Butz N, Vandamme TF. (2002). Pulmonary drug delivery systems: recent developments and prospects. *Crit Rev Ther Drug Carrier Syst*, 19:425–498.
58. Labiris NR, Dolovich MB. (2003). Pulmonary drug delivery. Part I: physiological factors affecting therapeutic effectiveness of aerosolized medications. *Br J Clin Pharmacol*, 56:588–599.
59. Byron PR. (1986). Prediction of drug residence times in regions of the human respiratory tract following aerosol inhalation. *J Pharm Sci*, 75:433–438.
60. Jaques PA, Kim CS. (2000). Measurement of total lung deposition of inhaled ultrafine particles in healthy men and women. *Inhal Toxicol*, 12:715–731.
61. Henning A, Hein S, Schneider M, Bur M, Lehr CM. (2010). Pulmonary drug delivery: medicines for inhalation. In: Schaefer-Korting M (Ed.) *Drug delivery*. Springer, Berlin, Heidelberg, Germany. 176.
62. Tsapis N, Bennett D, Jackson B, Weitz DA, Edwards DA. (2002). Trojan particles: large porous carriers of nanoparticles for drug delivery. *Proc Natl Acad Sci USA*, 99:12001–12005.
63. Sham JO, Zhang Y, Finlay WH, Roa WH, Löbenberg R. (2004). Formulation and characterization of spray-dried powders containing nanoparticles for aerosol delivery to the lung. *Int J Pharm*, 269:457–467.
64. Hadinoto K, Phanapavudhikul P, Kewu Z, Tan RBH. (2006). Novel formulation of large hollow nanoparticles aggregates as potential carriers in inhaled delivery of nanoparticulate drugs. *Ind Eng Chem Res* 45:3697–3706.

65. Hadinoto K, Zhu K, Tan RB. (2007). Drug release study of large hollow nanoparticulate aggregates carrier particles for pulmonary delivery. *Int J Pharm*, 341:195–206.
66. Hofmann W, Asgharian B. (2003). The effect of lung structure on mucociliary clearance and particle retention in human and rat lungs. *Toxicol Sci*, 73:448–456.
67. Schürch S, Gehr P, Im Hof V, Geiser M, Green F. (1990). Surfactant displaces particles toward the epithelium in airways and alveoli. *Respir Physiol*, 80:17–32.
68. Geiser M. (2010). Update on macrophage clearance of inhaled micro- and nanoparticles. *J Aerosol Med Pulm Drug Deliv*, 23:207–217.
69. Takenaka S, Karg E, Kreyling WG, Lentner B, Möller W, Behnke-Semmler M et al. (2006). Distribution pattern of inhaled ultrafine gold particles in the rat lung. *Inhal Toxicol*, 18:733–740.
70. Semmler-Behnke M, Takenaka S, Fertsch S, Wenk A, Seitz J, Mayer P et al. (2007). Efficient elimination of inhaled nanoparticles from the alveolar region: evidence for interstitial uptake and subsequent reentrainment onto airways epithelium. *Environ Health Perspect*, 115:728–733.
71. Geiser M, Casaulta M, Kupferschmid B, Schulz H, Semmler-Behnke M, Kreyling W. (2008). The role of macrophages in the clearance of inhaled ultrafine titanium dioxide particles. *Am J Respir Cell Mol Biol*, 38:371–376.
72. Hoeben BJ, Burgess DS, McConville JT, Najvar LK, Talbert RL, Peters JJ et al. (2006). *In vivo* efficacy of aerosolized nanostructured itraconazole formulations for prevention of invasive pulmonary aspergillosis. *Antimicrob Agents Chemother*, 50:1552–1554.
73. McConville JT, Overhoff KA, Sinswat P, Vaughn JM, Frei BL, Burgess DS et al. (2006). Targeted high lung concentrations of itraconazole using nebulized dispersions in a murine model. *Pharm Res*, 23:901–911.
74. Vaughn JM, McConville JT, Burgess D, Peters JJ, Johnston KP, Talbert RL et al. (2006). Single dose and multiple dose studies of itraconazole nanoparticles. *Eur J Pharm Biopharm*, 63:95–102.
75. Vaughn JM, Wiederhold NP, McConville JT, Coalson JJ, Talbert RL, Burgess DS et al. (2007). Murine airway histology and intracellular uptake of inhaled amorphous itraconazole. *Int J Pharm*, 338:219–224.
76. Garbuzenko OB, Saad M, Pozharov VP, Reuhl KR, Mainelis G, Minko T. (2010). Inhibition of lung tumor growth by complex pulmonary delivery of drugs with oligonucleotides as suppressors of cellular resistance. *Proc Natl Acad Sci USA*, 107:10737–10742.
77. Brigger I, Dubernet C, Couvreur P. (2002). Nanoparticles in cancer therapy and diagnosis. *Adv Drug Deliv Rev*, 54:631–651.
78. Hitzman CJ, Wattenberg LW, Wiedmann TS. (2006). Pharmacokinetics of 5-fluorouracil in the hamster following inhalation delivery of lipid-coated nanoparticles. *J Pharm Sci*, 95:1196–1211.
79. Azarmi S, Tao X, Chen H, Wang Z, Finlay WH, Löbenberg R et al. (2006). Formulation and cytotoxicity of doxorubicin nanoparticles carried by dry powder aerosol particles. *Int J Pharm*, 319:155–161.
80. Sharma S, White D, Imondi AR, Placke ME, Vail DM, Kris MG. (2001). Development of inhalational agents for oncologic use. *J Clin Oncol*, 19:1839–1847.
81. Otterson GA, Villalona-Calero MA, Sharma S, Kris MG, Imondi A, Gerber M et al. (2007). Phase I study of inhaled Doxorubicin for patients with metastatic tumors to the lungs. *Clin Cancer Res*, 13:1246–1252.
82. Kraft WK, Steiger B, Beussink D, Quiring JN, Fitzgerald N, Greenberg HE et al. (2004). The pharmacokinetics of nebulized nanocrystal budesonide suspension in healthy volunteers. *J Clin Pharmacol*, 44:67–72.
83. Sung JC, Pulliam BL, Edwards DA. (2007). Nanoparticles for drug delivery to the lungs. *Trends Biotechnol*, 25:563–570.
84. Chono S, Tanino T, Seki T, Morimoto K. (2007). Uptake characteristics of liposomes by rat alveolar macrophages: influence of particle size and surface mannose modification. *J Pharm Pharmacol*, 59:75–80.
85. du Toit LC, Pillay V, Danckwerts MP. (2006). Tuberculosis chemotherapy: current drug delivery approaches. *Respir Res*, 7:118.
86. Pandey R, Sharma A, Zahoor A, Sharma S, Khuller GK, Prasad B. (2003). Poly (DL-lactide-co-glycolide) nanoparticle-based inhalable sustained drug delivery system for experimental tuberculosis. *J Antimicrob Chemother*, 52:981–986.
87. Sharma A, Sharma S, Khuller GK. (2004). Lectin-functionalized poly (lactide-co-glycolide) nanoparticles as oral/aerosolized antitubercular drug carriers for treatment of tuberculosis. *J Antimicrob Chemother*, 54:761–766.
88. Pandey R, Khuller GK. (2005). Solid lipid particle-based inhalable sustained drug delivery system against experimental tuberculosis. *Tuberculosis (Edinb)*, 85:227–234.
89. Müller RH, Mäder K, Gohla S. (2000). Solid lipid nanoparticles (SLN) for controlled drug delivery - a review of the state of the art. *Eur J Pharm Biopharm*, 50:161–177.
90. Mansour HM, Rhee YS, Wu X. (2009). Nanomedicine in pulmonary delivery. *Int J Nanomedicine*, 4:299–319.
91. Pandey R, Khuller GK. (2005). Antitubercular inhaled therapy: opportunities, progress and challenges. *J Antimicrob Chemother*, 55:430–435.
92. Kawashima Y, Yamamoto H, Takeuchi H, Fujioka S, Hino T. (1999). Pulmonary delivery of insulin with nebulized DL-lactide/glycolide copolymer (PLGA) nanospheres to prolong hypoglycemic effect. *J Control Release*, 62:279–287.
93. Zhang Q, Shen Z, Nagai T. (2001). Prolonged hypoglycemic effect of insulin-loaded polybutylcyanoacrylate nanoparticles after pulmonary administration to normal rats. *Int J Pharm*, 218:75–80.
94. Renwick LC, Brown D, Clouter A, Donaldson K. (2004). Increased inflammation and altered macrophage chemotactic responses caused by two ultrafine particle types. *Occup Environ Med*, 61:442–447.
95. Oberdörster G. (1997). Pulmonary carcinogenicity of inhaled particles and the maximum tolerated dose. *Environ Health Perspect*, 105 Suppl 5:1347–1355.
96. Oberdörster G, Ferin J, Gelein R, Soderholm SC, Finkelstein J. (1992). Role of the alveolar macrophage in lung injury: studies with ultrafine particles. *Environ Health Perspect*, 97:193–199.
97. Renwick LC, Donaldson K, Clouter A. (2001). Impairment of alveolar macrophage phagocytosis by ultrafine particles. *Toxicol Appl Pharmacol*, 172:119–127.
98. Donaldson K, Stone V, Clouter A, Renwick L, MacNee W. (2001). Ultrafine particles. *Occup Environ Med*, 58:211–6, 199.
99. Nikula KJ. (2000). Rat lung tumors induced by exposure to selected poorly soluble nonfibrous particles. *Inhal Toxicol*, 12:97–119.
100. Borm PJ, Schins RP, Albrecht C. (2004). Inhaled particles and lung cancer, part B: paradigms and risk assessment. *Int J Cancer*, 110:3–14.
101. Driscoll KE, Deyo LC, Carter JM, Howard BW, Hassenbein DG, Bertram TA. (1997). Effects of particle exposure and particle-elicited inflammatory cells on mutation in rat alveolar epithelial cells. *Carcinogenesis*, 18:423–430.
102. Mühlfeld C, Gehr P, Rothen-Rutishauser B. (2008). Translocation and cellular entering mechanisms of nanoparticles in the respiratory tract. *Swiss Med Wkly*, 138:387–391.
103. Turco SJ. (2006). Intravenous admixtures. In: Troy BD (ed.) *Remington: the science and practice of pharmacy*. Lippincott Williams & Wilkins, Maryland, USA.
104. Shi Y, Porter W, Merdan T, Li LC. (2009). Recent advances in intravenous delivery of poorly water-soluble compounds. *Expert Opin Drug Deliv*, 6:1261–1282.
105. Akers, MJ. (2006). Parenteral preparations. In: Troy, BD (ed.) *Remington: the science and practice of pharmacy*. Lippincott Williams & Wilkins, Maryland, USA.
106. Yalkowsky SH, Krzyzaniak JF, Ward GH. (1998). Formulation-related problems associated with intravenous drug delivery. *J Pharm Sci*, 87:787–796.
107. Wong J, Brugger A, Khare A, Chaubal M, Papadopoulos P, Rabinow B et al. (2008). Suspensions for intravenous (IV) injection: a

- review of development, preclinical and clinical aspects. *Adv Drug Deliv Rev*, 60:939-954.
108. United State Pharmacopeia (USP). The National Formulary. (2007). Particulate matter in injections.
 109. Rabinow BE. (2004). Nanosuspensions in drug delivery. *Nat Rev Drug Discov*, 3:785-796.
 110. Haley B, Frenkel E. (2008). Nanoparticles for drug delivery in cancer treatment. *Urol Oncol*, 26:57-64.
 111. Storm G, Belliot SO, Daemen T, Lasic DD. (1995). Surface modification of nanoparticles to oppose uptake by the mononuclear phagocyte system. *Adv Drug Deliv Rev* 17:31-48.
 112. Cho K, Wang X, Nie S, Chen ZG, Shin DM. (2008). Therapeutic nanoparticles for drug delivery in cancer. *Clin Cancer Res*, 14:1310-1316.
 113. Davis ME, Chen ZG, Shin DM. (2008). Nanoparticle therapeutics: an emerging treatment modality for cancer. *Nat Rev Drug Discov*, 7:771-782.
 114. Farokhzad OC, Cheng J, Teply BA, Sherifi I, Jon S, Kantoff PW et al. (2006). Targeted nanoparticle-aptamer bioconjugates for cancer chemotherapy in vivo. *Proc Natl Acad Sci USA*, 103:6315-6320.
 115. Bartlett DW, Su H, Hildebrandt IJ, Weber WA, Davis ME. (2007). Impact of tumor-specific targeting on the biodistribution and efficacy of siRNA nanoparticles measured by multimodality *in vivo* imaging. *Proc Natl Acad Sci USA*, 104:15549-15554.
 116. Vauthier C, Dubernet C, Chauvierre C, Brigger I, Couvreur P. (2003). Drug delivery to resistant tumors: the potential of poly(alkyl cyanoacrylate) nanoparticles. *J Control Release*, 93:151-160.
 117. Schluep T, Hwang J, Cheng J, Heidel JD, Bartlett DW, Hollister B et al. (2006). Preclinical efficacy of the camptothecin-polymer conjugate IT-101 in multiple cancer models. *Clin Cancer Res*, 12:1606-1614.
 118. Ibrahim NK, Desai N, Legha S, Soon-Shiong P, Theriault RL, Rivera E et al. (2002). Phase I and pharmacokinetic study of ABI-007, a Cremophor-free, protein-stabilized, nanoparticle formulation of paclitaxel. *Clin Cancer Res*, 8:1038-1044.
 119. Desai NP, Soon-Shiong P, Yang A. (2007). Novel formulations of pharmacological agents, methods for the preparation thereof and methods for the use thereof. US Patent No. 0092563.
 120. Desai NP, Selvaraj R, Yang A, Soon-Shiong P. (2010). Compositions comprising poorly water soluble pharmaceutical agents and antimicrobial agents. US Patent No. 7771751.
 121. Desai N, Trieu V, Damascelli B, Soon-Shiong P. (2009). SPARC Expression Correlates with Tumor Response to Albumin-Bound Paclitaxel in Head and Neck Cancer Patients. *Transl Oncol*, 2:59-64.
 122. Desai N, Trieu V, Yao Z, Louie L, Ci S, Yang A et al. (2006). Increased antitumor activity, intratumor paclitaxel concentrations, and endothelial cell transport of cremophor-free, albumin-bound paclitaxel, ABI-007, compared with cremophor-based paclitaxel. *Clin Cancer Res*, 12:1317-1324.
 123. Cortes J, Saura C. (2010). Nanoparticle albumin-bound (nab (TM))-paclitaxel: improving efficacy and tolerability by targeted drug delivery in metastatic breast cancer. *EJC Suppl* 8:1-10.
 124. ten Tije AJ, Verweij J, Loos WJ, Sparreboom A. (2003). Pharmacological effects of formulation vehicles: implications for cancer chemotherapy. *Clin Pharmacokinet*, 42:665-685.
 125. Gradishar WJ, Tjulandin S, Davidson N, Shaw H, Desai N, Bhar P et al. (2005). Phase III trial of nanoparticle albumin-bound paclitaxel compared with polyethylated castor oil-based paclitaxel in women with breast cancer. *J Clin Oncol*, 23:7794-7803.